Date Submitted: Aug. 20, 2007
Protocol Title: Immunostaining for neprilysin
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Name of Lab PI: Takaomi C. Saido, Ph.D.
_X_. This submission has been approved by Lab PI

Categories: Place an X next to all that apply to this protocol. Please add a new category name if needed.

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<tr>
<th>Proteins</th>
<th>Methods</th>
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<td>Abeta</td>
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<td>Collection of Biofluids</td>
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Overview (Text that describes the purpose of the protocol)

This is a protocol for the immunostaining of neprilysin using the high temperature antigen unmasking technique and the immunoperoxidase-indirect or fluorescence-indirect tyramide signal amplification (TSA) method. Among various methods for antigen retrieval, including autoclaving, microwaving, formic acid or detergent treatment, and tryptic digestion, autoclaving at 121°C for 5 min gives the best results. Without autoclaving, no immunoreactivity is detected. Signals detected by this protocol are highly specific for neprilysin, because no signal is detected in the brain of neprilysin-deficient mice. For fixation of tissue specimen, 4% paraformaldehyde is preferable. A paraffin section is much better than frozen sections because of a procedure for autoclaving.
Reagents (List of reagents used in the protocol; include company and catalog number for antibodies so that we can link to our Antibody database)

- Primary antibody (anti-neprilysin): clone 56C6 (NCL-CD10-270, 1ml, Novocastra Lab.)
- Avidin/biotin blocking kit (SP-2001, Vector Lab.)
- Second antibody: Biotin conjugated-goat anti-mouse IgG$_1$ (1070-08, 1mg/ml, Southern Biotech.)
- TSA Biotin System (TSA-Indirect kit) (NEL-700A, NEN)
- Alexa Fluor 488-conjugated streptavidin (S-11223, Molecular Probe)
- Perma Fluor Aqueous Mountant (434990, Immunon, Pittsburgh, PA)
- 10mM Sodium citrate buffer, pH6.0.
- 0.3% H$_2$O$_2$ in methanol solution
- TN buffer: 0.1M Tris-HCl, 0.15M NaCl, pH 7.5.
- TNT wash buffer: 0.1M Tris-HCl, 0.15M NaCl, 0.05% Tween 20, pH 7.5.
- TNB (blocking buffer): 0.1M Tris-HCl, 0.15M NaCl, 0.5% blocking reagent, pH 7.5.
- ImmunoPure Metal Enhanced DAB substrate (#1850090, PIERCE)

Equipment (List of equipment needed for this protocol)

Confocal laser microscope/Fluorescence microscope

Protocol Procedure (Text that lists steps of protocol; may include images or video)

Immunofluorescence staining

See the manufacturer’s instructions for TSA biotin system and avidin/biotin blocking kit before starting!

1. Deparaffinize & rehydrate.
2. Autoclave in 10mM Sodium citrate buffer (pH6.0) at 121°C for epitope retrieval of antigen.
   *Don’t cap a stainless steel cooker containing slides!*
3. Rinse sections in tap water.
4. 0.3% H$_2$O$_2$ in methanol
5. Rinse in tap water.
6. Wash in TN.
7. Avidin blocking (Vector kit)
8. Biotin blocking (Vector kit)
9. TSA kit blocking
10. Primary antibody (x100-x200, diluted with TNB)

5 min
30 min
5 min
5 min x 3
15 min
15 min
30 min
4 °C o/n
11 Wash in TNT.  
12 Biotinylated second antibody (x2,000-3,000, diluted with TNB)  
13 Wash in TNT.  
14 Streptavidin-HRP (x100) (TSA-Indirect kit)  
15 Wash in TNT.  
16 Biotinyl tyramide amplification reagent (x50) (TSA-Indirect kit)  
17 Wash in TNT  
18 Streptavidin-Alexa488 (x500)  
19 Coverslip with Perma Fluor Aqueous Mountant to retard fading.  
20 Capture images with a microscope incorporating a confocal laser scanning system.

<table>
<thead>
<tr>
<th>Step</th>
<th>Description</th>
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<tbody>
<tr>
<td>11</td>
<td>Wash in TNT.</td>
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<tr>
<td>12</td>
<td>Biotinylated second antibody (x2,000-3,000, diluted with TNB)</td>
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<td>Capture images with a microscope incorporating a confocal laser scanning system.</td>
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**Peroxidase-DAB(Diaminobenzidine)- staining**

*See the PIERCE’s instructions for metal enhanced DAB substrate.*

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<tr>
<td>1-17</td>
<td>the same as above</td>
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</table>
| 18’  | Streptavidin-HRP (x100) (TSA-Indirect kit)  
19’  | Wash in TNT.  
20’  | DAB |

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**References** (List up to 5 key citations that describe applications of the protocol)

